

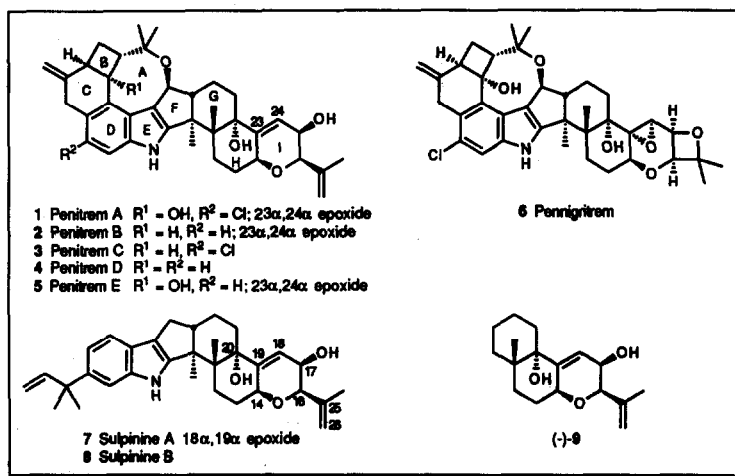
## AN END-GAME STRATEGY FOR CONSTRUCTION OF THE G-H-I RINGS OF PENITREM D, A TREMORGENIC INDOLE ALKALOID

Amos B. Smith, III,\* Mitsuki Ohta, William M. Clark, and James W. Leahy

Department of Chemistry, the Laboratory for Research on the Structure of Matter, and the Monell Chemical Senses Center,  
University of Pennsylvania, Philadelphia, Pennsylvania 19104, U.S.A.

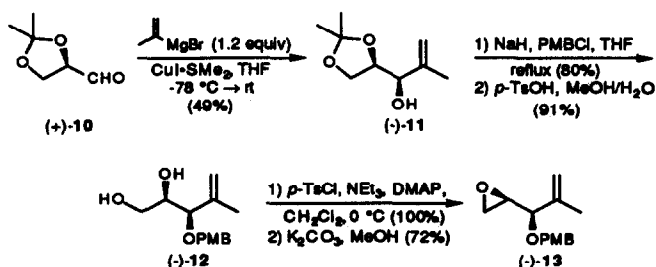
**Summary:** A synthetic approach to the G-H-I ring system of penitrem D has been developed in a model study. To generate the requisite carbon framework, the Stork protocol was utilized for coupling of a scalemic metaloenamine with an epoxide derived from D-glyceraldehyde. Pyran ring formation was then effected via  $\text{BF}_3 \cdot \text{OEt}_2$ -promoted cyclization of a  $\delta$ -hydroxy allylic acetate.

The tremorgenic indole alkaloids embody unusual, architecturally complex polycyclic skeleta and also present distinctive biological profiles, inducing tremors in animals and possessing significant insecticidal activity.<sup>1,2</sup> Prominent members of the class include the penitrem tremorgenic mycotoxins (1-6), first isolated from *Penicillium crustosum*,<sup>3</sup> which cause severe neurological dysfunction in livestock, not unlike the symptoms observed in human disorders such as Wilson's disease and multiple sclerosis.<sup>3a,4,5</sup> Sulpinine A and B (7 and 8),<sup>6</sup> recently isolated from the sclerotia of *Aspergillus sulphureus*, share the terminal G-H-I ring substructures and 7 exhibits pronounced cytotoxicity toward human lung, breast, and colon carcinomas.<sup>6</sup> Our long-standing interest in the construction of the indole alkaloid tremorgens<sup>7</sup> has led to a penitrem D (4) synthetic venture.<sup>8</sup> Herein we describe an end-game strategy for penitrem D whereby we have prepared (-)-9, a model compound comprising the G-H-I ring system of 4.



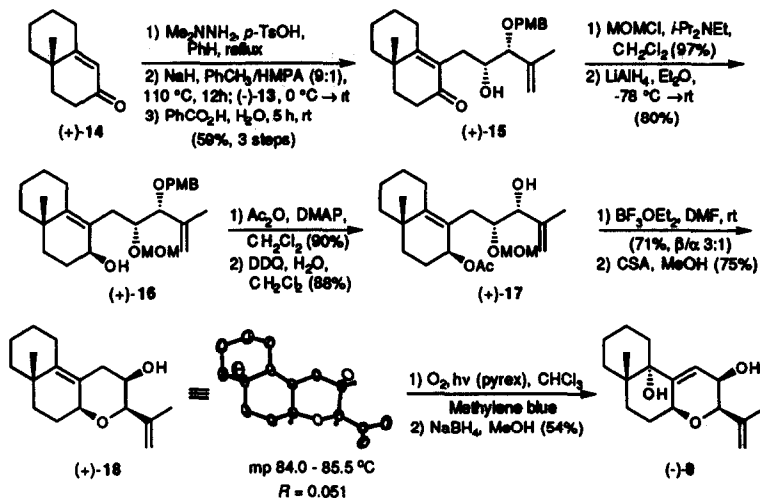
Wishing to exploit and extend an approach developed earlier,<sup>7</sup> we anticipated the use of a metaloenamine<sup>9</sup> as a nucleophilic H-ring synthon. Accordingly, we designed epoxide 13<sup>10</sup> to serve as an electrophilic progenitor of the pyran ring. Addition of the cuprate prepared from isopropenylmagnesium bromide to the acetonide derivative of D-glyceraldehyde [(+)-10]<sup>11</sup> gave a 95:5 mixture of diastereomers (Scheme 1). After purification of (-)-11<sup>12,13</sup> by distillation, hydroxyl protection as the PMB ether and deketalization smoothly generated diol (-)-12.<sup>13</sup> The epoxide (-)-13<sup>13</sup> was then formed via tosylation of the primary alcohol and cyclization with potassium carbonate in methanol.

## Scheme 1



With (-)-13 in hand, we turned to the metalloenamine coupling with octalone (+)-14<sup>14</sup> (Scheme 2). Following the procedure of Stork,<sup>9</sup> the dimethyl hydrazone of (+)-14 was deprotonated with sodium hydride in toluene/HMPA (9:1) at 110 °C. Addition of the epoxide (-)-13 at 0 °C and careful hydrolysis<sup>15</sup> of the hydrazone then afforded (+)-15<sup>13</sup> in 59% overall yield. Previously we found that stringent deoxygenation of the reaction mixture was essential for successful coupling,<sup>7h</sup> but in this instance such precaution was not necessary. The hydroxyl group in (+)-15 was next converted to the corresponding MOM ether and 1,2-reduction of the resultant ketone was attempted. Initially we intended to generate the  $\alpha$  alcohol, permitting direct conversion to the desired pyran. In the event, however, a variety of reduction protocols [LAH,  $\beta/\alpha$  9:1; LAH/AIMe<sub>3</sub>, 2.75:1; DIBAL-H, 1.3:1; DIBAL-H/AIMe<sub>3</sub>, 3:1] furnished predominantly the  $\beta$  alcohol (+)-16.<sup>13</sup>

## Scheme 2



To take advantage of the observed selectivity, we sought to induce cyclization with double inversion [i.e., net retention of configuration at C(14)]; one attractive tactic involved the intermediacy of a  $\pi$ -allyl palladium complex.<sup>16</sup> To this end, acetylation of (+)-16 and oxidative cleavage of the PMB ether with DDQ provided (+)-17<sup>13</sup> in excellent yield [80% for two steps, Scheme 2]. Unfortunately the cyclization required stringent conditions [(PPh<sub>3</sub>)<sub>2</sub>PdCl<sub>2</sub> (0.2 equiv), (*t*-Pr)<sub>2</sub>NEt (1.0 equiv), DMF, 150 °C, 12 h] and furnished the desired pyran in low yield with poor stereoselectivity (30-40%,  $\beta/\alpha$  1.5:1). Upon further investigation, we discovered that Lewis acid-promoted cyclization of the allylic acetate in polar solvents led to significant improvements in both yield and selectivity. For example, exposure of (+)-17 to BF<sub>3</sub>·OEt<sub>2</sub> (0.5 equiv) in DMF at

room temperature generated the pyran as a 3:1 mixture of  $\beta$  and  $\alpha$  epimers in 71% yield;  $\text{TiCl}_4$  (0.5 equiv) in DMF resulted in higher selectivity (5:1) but a lower yield (42%). Interestingly, this approach afforded similar yields and diastereomer ratios upon cyclization of the  $\alpha$  epimer of allylic acetate (+)-17. Detailed studies of these reactions are in progress.

Following the successful elaboration of the pyran ring, completion of the synthesis of **9** entailed introduction of the C(20) hydroxyl substituent with olefin transposition as well as removal of the MOM ether (Scheme 2). The latter was effected with CSA in methanol; at this point the diastereomers were easily separable by flash chromatography, and the stereochemistry of (+)-18<sup>13</sup> was confirmed through the aegis of single-crystal X-ray analysis.<sup>17</sup> Oxidation of (+)-18 with  $^1\text{O}_2$  (methylene blue sensitizer,  $\text{CHCl}_3$ ) followed by  $\text{NaBH}_4$  reduction of the intermediate hydroperoxide then afforded (-)-**9** as the sole product. As shown in Table 1, the stereochemical assignment at C(20) is strongly supported by comparison with the  $^1\text{H}$  and  $^{13}\text{C}$  NMR data for sulpinine B (**8**). Our efforts to employ this end-game strategy in the total synthesis of penitrem D will be reported in due course.

Table 1.  $^1\text{H}$  and  $^{13}\text{C}$  NMR Comparisons of Sulpinine B (**8**) and (-)-**9**.<sup>a</sup>

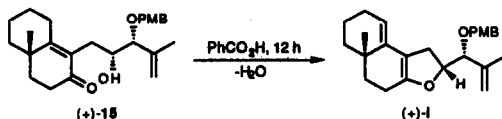
Carbon	Sulpinine B ( <b>8</b> )		(-)- <b>9</b>	
	$^1\text{H}$	$^{13}\text{C}$	$^1\text{H}$	$^{13}\text{C}$
14	4.61 (br dd, 9.1, 8.8 Hz)	73.7	4.52 (dd, 11.0, 6.9 Hz)	74.1
16	3.86 (br s)	79.1	3.87 (br s)	78.8
17	3.96 (br d, 5.6 Hz)	62.8	3.95 (dd, 5.8, 1.7 Hz)	62.9
18	5.82 (br d, 4.7 Hz)	118.7	5.82 (dd, 5.8, 1.7 Hz)	119.8
19	—	148.1	—	147.6
20	—	77.7	—	75.1
25	—	141.6	—	141.8
26	5.20 (br s) 5.03 (br s)	111.7	5.17 (br s) 5.02 (br s)	111.5

<sup>a</sup>Chemical shifts are reported in ppm. Data reported for sulpinine B<sup>6</sup> were recorded in  $\text{CDCl}_3$  at 600 and 75.6 MHz. Data for (-)-**9** were recorded in  $\text{CDCl}_3$  at 500 and 125.7 MHz.

**Acknowledgment.** The financial support of the National Institutes of Health (Institute of Neurology, Communicative Disorders and Stroke) through grant 18254 is gratefully acknowledged. We also thank Drs. George Furst and Patrick J. Carroll, and Mr. John Dykins, of the University of Pennsylvania Spectroscopic Service Centers, for assistance in securing and interpreting high-field NMR spectra, X-ray crystal structures, and mass spectra, respectively.

## References and Notes

1. (a) Gallagher, R. T.; White, E. P.; Mortimer, P. H. *N. Z. Vet. J.* **1981**, *29*, 189. (b) Cole, R. J.; Dornier, J. W.; Lansden, J. A.; Cox, R. H.; Pape, C.; Cunfer, B.; Nicholson, S. S.; Bedell, D. M. *J. Agric. Food Chem.* **1977**, *25*, 1197.
2. (a) Dowd, P. F.; Cole, R. J.; Vesonder, R. F. *J. Antibiot.* **1988**, *41*, 1868. (b) Prestridge, R. A.; Gallagher, R. T. *Proc. N. Z. Weed Pest Control Conf.* **1985**, *38*, 38. (c) Prestridge, R. A.; Gallagher, R. T. *Ecol. Entomol.* **1988**, *13*, 429.
3. For structure elucidation studies, see: (a) De Jesus, A. E.; Steyn, P. S.; Van Heerden, F. R.; Vleggar, R.; Wessels, P. L.; Hull, W. E. *J. Chem. Soc., Chem. Commun.* **1981**, 289. (b) De Jesus, A. E.; Hull, W. E.; Steyn, P. S.; Van Heerden, F. R.; Vleggar, R.; Wessels, P. L. *J. Chem. Soc., Chem. Commun.* **1982**, 837. (c) De Jesus, A. E.; Steyn, P. S.; Van Heerden, F. R.; Vleggar, R.; Wessels, P. L.; Hull, W. E. *J. Chem. Soc., Perkin Trans. 1* **1983**, 1847. (d) De Jesus, A. E.; Gorst-Allman, C. P.; Steyn, P. S.; Van Heerden, F. R.; Vleggar, R.; Wessels, P. L.; Hull, W. E. *J. Chem. Soc., Perkin Trans. 1* **1983**, 1863. For isolation studies, see: (e) Wilson, B. J.; Wilson, C. H.; Hayes, A. W. *Nature* **1968**, *220*, 77. (f) Ciegler, A. *Appl. Microbiol.* **1969**, *18*, 128. (g) Ciegler, A.; Pitt, J. I. *Mycopathol. Mycol. Appl.* **1970**, *42*, 119. (h) Hou, C. T.; Ciegler, A.; Hesseltine, C. W. *Can. J. Microbiol.* **1971**, *17*, 599. (i) Pitt, J. I. *Mycologia* **1979**, *71*, 1166. (j) Wagener, R. E.; Davis, N. D.; Diener, U. L. *Appl. Environ. Microbiol.* **1980**, *39*, 882. (k) Vesonder, R. F.; Tjarks, L.; Rohwedder, W.; Kieswetter, D. O. *Experientia* **1980**, *36*, 1308. (l) Kyriakidis, N.; Waight, E. S.; Day, J. B.; Mantle, P. G. *Appl. Environ. Microbiol.* **1981**, *42*, 61.
4. (a) Peterson, D. W.; Penny, R. H. C.; Day, J. B.; Mantle, P. G. *Res. Vet. Sci.* **1977**, *33*, 183. (b) Hayes, A. W.; Hood, R. D. *Toxicol.* **1978**, *16*, 92. (c) Hayes, A. W.; Presley, D. B.; Neville, J. A. *Toxicol. Appl. Pharmacol.* **1978**, *35*, 311. (d) Hayes, A. W.; Phillips, R. D.; Wallace, L. C. *Toxicol.* **1977**, *15*, 293. (e) Sobotka, T. J.; Brodie, R. E.; Spald, S. L. *Pharmacol.* **1978**, *16*, 287. (f) Mantle, P. G.; Mortimer, P. H.; White, E. P. *Res. Vet. Sci.* **1978**, *24*, 49.
5. Penn, J.; Biddle, J. R.; Mantle, P. G.; Bilton, J. N.; Sheppard, R. N. *J. Chem. Soc., Perkin Trans. 1* **1992**, 23.
6. Laakso, J. A.; Gloer, J. B.; Wicklow, D. T.; Dowd, P. F. *J. Org. Chem.* **1992**, *57*, 2066.
7. (a) Smith, A. B., III; Mewshaw, R. *J. Am. Chem. Soc.* **1985**, *107*, 1769. (b) Smith, A. B., III; Visnick, M. *Tetrahedron Lett.* **1985**, *26*, 3757. (c) Smith, A. B., III; Visnick, M.; Haseltine, J. N.; Sprengeler, P. A. *Tetrahedron* **1986**, *42*, 2957. (d) Smith, A. B., III; Leenay, T. L. *Tetrahedron Lett.* **1988**, *29*, 2787. (e) Smith, A. B., III; Leenay, T. L. *Tetrahedron Lett.* **1988**, *29*, 2791. (f) Mewshaw, R. E.; Taylor, M. D.; Smith, A. B., III. *J. Org. Chem.* **1989**, *54*, 3449. (g) Smith, A. B., III; Leenay, T. L. *J. Am. Chem. Soc.* **1989**, *111*, 5761. (h) Smith, A. B., III; Sunazuka, T.; Leenay, T. L.; Kingery-Wood, J. *J. Am. Chem. Soc.* **1990**, *112*, 8197. (i) Smith, A. B., III; Kingery-Wood, J.; Leenay, T. L.; Nolen, E. G.; Sunazuka, T. *J. Am. Chem. Soc.* **1992**, *114*, 1438.
8. For previous synthetic studies on penitrem D, see: (a) Haseltine, J. N.; Visnick, M.; Smith, A. B., III. *J. Org. Chem.* **1988**, *53*, 6160; (b) Smith, A. B., III; Haseltine, J. N.; Visnick, M. *Tetrahedron* **1989**, *45*, 2431.
9. Stork, G.; Benaim, J. *J. Am. Chem. Soc.* **1971**, *93*, 5938. Also see: Stork, G.; Benaim, J. *Org. Synth.* **1977**, *57*, 69.
10. For related derivatives, see: Sato, F.; Kobayashi, Y.; Takahashi, O.; Chiba, T.; Takeda, Y.; Kusakabe, M. *J. Chem. Soc., Chem. Commun.* **1985**, 1636.
11. Jackson, D. Y. *Synth. Commun.* **1988**, *18*, 337.
12. A preparation of this compound has recently appeared: Lee, H. W.; Lee, I.-Y. *C. Synlett* **1991**, 871.
13. The structure assigned to each new compound is in accord with its infrared, 500-MHz <sup>1</sup>H NMR, and 125-MHz <sup>13</sup>C NMR spectra, as well as appropriate parent ion identification by high resolution mass spectrometry. In addition, analytical samples of **12**, **16**, and **17** gave satisfactory C and H combustion analysis.
14. Pfau, M.; Revial, G.; Guingant, A.; d'Angelo, J. *J. Am. Chem. Soc.* **1985**, *107*, 273.
15. Overexposure of (+)-**15** to the hydrolysis conditions resulted in extensive dehydration, furnishing (+)-**1**.



16. Heck, R. F. *Palladium Reagents in Organic Synthesis*; Academic Press: London, 1985; Chapter 5.
17. Carroll, P. J., University of Pennsylvania, unpublished results.